CERTIFICATE

This certificate is issued in support of an application for Patent registration in a country outside New Zealand pursuant to the Patents Act 1953 and the Regulations thereunder.

I hereby certify that annexed is a true copy of the Provisional Specification as filed on 31 May 2004 with an application for Letters Patent number 533245 made by THE MALAGHAN INSTITUTE OF MEDICAL RESEARCH; UNIVERSITY OF OTAGO and AGRESEARCH LIMITED.
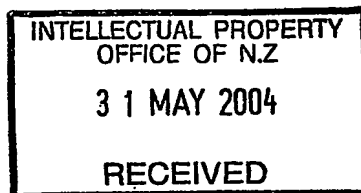
Dated 11 January 2005.

Neville Harris
Commissioner of Patents, Trade Marks and Designs

533245

NEW ZEALAND

PATENTS ACT, 1953

PROVISIONAL SPECIFICATION

SYNTHETIC MOLECULES HAVING IMMUNE ACTIVITY

We, **THE MALAGHAN INSTITUTE OF MEDICAL RESEARCH**, a New Zealand non-profit organisation of Mein Street, Newtown, Wellington, New Zealand; and **UNIVERSITY OF OTAGO**, a body corporate established under the University of Otago ordinance 1869, of Leith Street, Dunedin, New Zealand; and **AGRESEARCH LIMITED**, a New Zealand company and Crown Research Institute under the Crown Research Institutes Act 1992, of 5th Floor, Tower Block, Ruakura Research Centre, East Street, Hamilton, New Zealand, do hereby declare this invention to be described in the following statement:

FIELD OF INVENTION

The present invention relates to synthetic molecules having biological activity, in particular immune activity including PIM or PIM-like activity, specifically, although by no means exclusively, for use as an immune system modifier.

5

BACKGROUND

PIM (acyl glyceryl phosphatidylinositol manno-oligosaccharide) is an immunogenic component of mycobacterial cell walls which is capable of treating or preventing inflammatory or immune cell-mediated diseases and disorders such as asthma, allergic
10 rhinitis, dermatitis, psoriasis, inflammatory bowel disease including Crohn's disease and ulcerative colitis, rheumatoid arthritis, multiple sclerosis, diabetes, systemic lupus erythematosis and atherosclerosis.

WO 02/02140 discloses an immunogenic composition comprising PIM which is effective in
15 the treatment and prevention of Th2 mediated disease, particularly asthma. In particular, the PIM vaccine appears to act by suppressing the allergic response which would normally cause a recruitment and activation of eosinphils to the lung causing chronic swelling and inflammation of the airways that affects the breathing of sufferers. Experiments using a mouse model of airway eosinophilia illustrated that administration of the PIM composition
20 resulted in a dose dependent decrease in the number of eosinophils in the lungs of such mice.

Currently a heterogeneous mixture of PIM species is produced by isolating the PIM fraction from dead mycobacterial organisms using a series of chemical purification steps as disclosed in WO 02/02140 and in Severn et al, 1997. This purification process is laborious and not
25 suitable for large scale manufacture of PIM.

In particular, the PIM fraction can be contaminated by lipopolysaccharides such as endotoxins which are also known to induce an immunological response and therefore may mask or interfere with the biological activity of such a PIM extract.

30

95896-1

PIM exists in nature in many different forms. For example the number of mannose and acyl residues may vary. Different acyl forms have been purified using sophisticated analytical tools such as MALDI-MS and two-dimensional NMR. (Gilleron *et al* 2001; Gilleron *et al* 2003). In particular native PIM_2 and PIM_6 have been purified, characterised and their biological activity demonstrated in that these compounds stimulate macrophages to produce cytokines.

In addition, a number of glycophospholipid compounds have been synthesised and either polymerised to form synthetic cell membranes, bilayers, films, liposomes for use in drug delivery systems (US 6,071,532; US 6,171,614; JP 06-271597) or used in therapeutic compositions for treating inflammatory disorders (US 2002/0028823; US 6024940; US 2002/0032195; US 2003/0008848; US 2003/0022913).
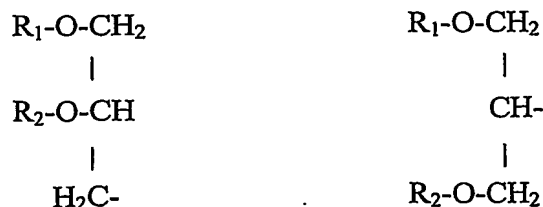
It is an object of the present invention to provide novel synthetic molecules having biological activity, including PIM or PIM-like activity, which may be synthesised using highly efficient and economical chemical processes suitable for manufacture on a large scale and free of endotoxin and/or to provide the public with a useful choice.

SUMMARY OF INVENTION

According to the present invention there is provided a synthetic molecule of formula I:

$$A\text{-}B\text{-}E\text{-}D \hspace{4cm} (I)$$

wherein represents R, or a glyceride group having the formula Ia or Ib:

$$R_1\text{-}O\text{-}CH_2 \hspace{5cm} R_1\text{-}O\text{-}CH_2$$
$$| \hspace{7cm} |$$
$$R_2\text{-}O\text{-}CH \hspace{5.5cm} CH\text{-}$$
$$| \hspace{7cm} |$$
$$H_2C\text{-} \hspace{5.5cm} R_2\text{-}O\text{-}CH_2$$

(Ia) \hspace{5cm} (Ib)

wherein R is H or a linear or branched alkyl of up to 40 carbon atoms, preferably 6-22, more preferably 10-20, and most preferably 16-20 carbon atoms; R_1 and R_2 are independently H, alkyl or acyl and wherein the alkyl or acyl groups are linear or branched having up to 40 carbon atoms, preferably 6-22, more preferably 10-20, and most preferably 16-20 carbon atoms;

B is selected from the group comprising phosphate, phosphonate, sulfonate, carbamate, and phosphothionate;

E comprises a spacer or linker group providing a linkage between groups B and D and may be selected from $(CH_2)_n$; $-(CH_2)_2-(O-CH_2-CH_2)_n-$; -cyclohexyl-; and $-CHR_3-CHR_4-$ wherein R_3 and R_4 are independently H, CH_2OH, CH_2-, or $(CH(OH))_n-CH_2OH$ or

$CH((CHOH)_nCH_2OH)-$; and wherein n=1 to 40;

D comprises at least one sugar moiety selected from the group comprising D-mannose, D-galactose, D-glucose, D-glucosamine, N-acetylglucosamine, and 6-deoxy-L-mannose, wherein when D is more than one sugar moiety, the sugar moiety may comprise a single chain of the same or different sugar moieties, or may comprise two or more separate sugar moieties or chains of sugar moieties attached to E at different sites;

with the proviso that when E is $-(CH_2)_n-$ wherein n=2 to 16, B is phosphate and D is a monosaccharide or an oligosaccharide, R_1 and R_2 of A are not both alkyl.

Preferably, D comprises a monosaccharide or oligosaccharide chain of 2 to 12, more preferably 2 to 6, α-1,2 and/or α-1,6 linked sugar moieties which are O-linked to carbon atoms on spacer group E. More preferably, D comprises one or more monosaccharide or oligosaccharide chains of 2 to 6 sugar moieties. One or more of the sugar moieties D may be acylated.

Typically, R_1 and R_2 are fatty acids independently selected from the group comprising myristate, palmitate, heptadecanoate, stearate, tuberculostearate or linolenate; B is phosphate;

E is $-CHR_3CHR_4-$, wherein R_3 is CH_2- and R_4 is H; and D is at least one sugar moiety comprising D-mannose or oligosaccharide chain of α-1,2 and/or α-1,6-linked mannose residues.

5 In another embodiment the present invention provides a pharmaceutical composition comprising at least one compound of formula (I) or a pharmaceutically acceptable salt thereof and a pharmaceutically acceptable carrier.

In a further embodiment the present invention provides a use of a compound of formula (I) or
10 a pharmaceutically acceptable salt thereof in the manufacture of a medicament for treating or preventing an inflammatory or immune cell-mediated diseases or disorders, such as asthma, allergic rhinitis, dermatitis, psoriasis, inflammatory bowel disease including Crohn's disease and ulcerative colitis, rheumatoid arthritis, multiple sclerosis, diabetes, systemic lupus erythmatosis and atherosclerosis.

15

The present invention further provides a use of a compound of formula (I) or a pharmaceutically acceptable salt thereof in the manufacture of an adjuvant for use in enhancing the immune response to an antigen. In addition, the invention provides an adjuvant composition comprising an effective adjuvanting amount of a compound of formula (I) or a
20 pharmaceutically acceptable salt thereof.

In a further embodiment the present invention provides a method of treating or preventing an inflammatory or immune cell-mediated disease or disorder comprising administering an effective amount of a compound of formula (I) or a pharmaceutically acceptable salt thereof
25 to a patient in need thereof. Typically, the patient is a human patient. Typically, the inflammatory or immune cell-mediated disease or disorder is asthma, allergic rhinitis, dermatitis, psoriasis, inflammatory bowel disease including Crohn's disease and ulcerative colitis, rheumatoid arthritis, multiple sclerosis, diabetes, systemic lupus erythmatosis and atherosclerosis.

30

95896-1

In a further embodiment the present invention provides a process for preparing synthetic molecules of formula I.

DESCRIPTION OF THE FIGURES

5   The invention will now be described by reference to the figures of the accompanying drawings in which:

Figure 1 shows a schematic representation of the synthesis of a compound of the invention named Compound 7;

10

Figure 2 shows a schematic representation of the synthesis of a compound of the invention named Compound 15;

Figure 3 shows a schematic representation of the synthesis of a comparative compound

15   named Compound 17; and

Figure 4 shows the mean (± s.e.m.) eosinophil count (x 10^6) after administration of a compound of the present invention and a comparative PIM extract using the mouse *in vivo* induced airway eosinophilia model.

20

DETAILED DESCRIPTION OF THE INVENTION

As broadly outlined above, the present invention is directed to novel synthetic molecules having biological activity, including PIM or PIM-like activity, which are useful in treating an inflammatory or immune cell-mediated disease or disorder in a patient, and in particular in the

25   treatment of asthma in an asthmatic and/or for reducing the risk of developing airway eosinophilia and thus asthma in a non-asthmatic in much the same way as natural PIM has been reported (WO 02/02140). In addition, it is expected that the synthetic molecules of the present invention will be useful in treating allergic rhinitis, dermatitis, psoriasis, inflammatory

bowel disease including Crohn's disease and ulcerative colitis, rheumatoid arthritis, multiple sclerosis, diabetes, systemic lupus erythmatosis and atherosclerosis.

5   The structure of the natural PIM molecule, isolated, for example, from a mycobacterium is made up of a diacylglycerol unit, a phosphate group, C-2 and C-6 mannopyranose units and an inositol unit as follows:

10
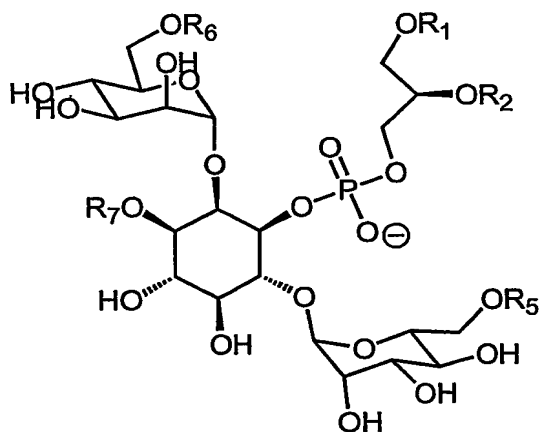
where R_1, R_2, R_6 and R_7 are independently either hydrogen or an acyl group selected from palmitate, stearate and tuberculostearate; and R_5 is either hydrogen or a monooligosaccharide.
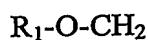
It is not known which part of the natural PIM molecule is responsible for its
15   immunomodulating effects, although a deacylated natural PIM has been shown to be incapable of eliciting an immune response (WO 02/02140).

The present invention provides synthetic molecules which have similar or enhanced immunomodulating activity compared to natural PIM.
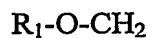
20

The molecules of the present invention may be synthesised using known methods as described in the Examples below. Specifically, the synthetic molecules of the present invention comprise a compound of the formula I:

A-B-E-D   (I)

95896-1

wherein A represents R or a glyceride group having the formula Ia or Ib:

R_1-O-CH$_2$ R_1-O-CH$_2$
    | |

5   R_2-O-CH   CH-
       | |
     H$_2$C- R_2-O-CH$_2$

       (Ia)   (Ib)

10

wherein R is H or a linear or branched alkyl of up to 40 carbon atoms preferably 6-22, more preferably 10-20, and most preferably 16-20 carbon atoms; R_1 and R_2 are independently H, alkyl or acyl and wherein the alkyl or acyl groups are linear or branched having up to 40 carbon atoms, preferably 6-22, more preferably 10-20, and most preferably 16-20 carbon

15   atoms;

B is selected from the group comprising phosphate, phosphonate, sulfonate, carbamate, and phosphothionate;

20   E comprises a spacer or linker group providing a linkage between groups B and D and may be selected from -(CH$_2$)$_n$-; -(CH$_2$)$_2$-(O-CH$_2$-CH$_2$)$_n$-; -cyclohexyl-; and -CHR$_3$-CHR$_4$- wherein R_3 and R_4 are independently H, CH$_2$OH, CH$_2$- or (CH(OH))$_n$-CH$_2$OH or CH((CHOH)$_n$CH$_2$OH)-; and wherein n=1 to 40;

25   D comprises at least one sugar moiety selected from the group comprising D-mannose, D-galactose, D-glucose, D-glucosamine, N-acetylglucosamine and 6-deoxy-L-mannose, wherein when D is more than one sugar moiety, the sugar moiety may comprise a single chain of the same or different sugar moieties, or may comprise two or more separate sugar moieties or chains of sugar moieties attached to E at different sites;

30

with the proviso that when E is -(CH$_2$)$_n$- wherein n=2 to 16, B is phosphate and D is a monosaccharide or oligosaccharide, R_1 and R_2 of A are not both alkyl.

Preferably, D comprises a monosaccharide or an oligosaccharide chain of 2 to 12, more preferably 2 to 6, α-1,2 and/or α-1,6 linked sugar moieties which are O-linked to carbon atoms on spacer group E. More preferably, D comprises one or more monosaccharides or oligosaccharide chains of 2 to 6 sugar moieties. One or more of the sugar moieties of D may be acylated.

Typically, R_1 and R_2 are fatty acids independently selected from the group comprising myristate, palmitate, heptadecanoate, stearate, tuberculostearate or linolenate; B is phosphate; E is $-CHR_3CHR_4-$ where R_3 is CH_2- and R_4 is H; and D is at least one sugar moiety comprising D-mannose or oligosaccharide chain of α-1,2- and/or α-1,6- linked mannose residues.

Compounds where R, R_1 and/or R_2 comprise long chain acyl or alkyl of up to 60 carbon atoms are contemplated and may be synthesised, although synthesis may be expensive and/or difficult as would be appreciated by a skilled worker. Such long acyl/alkyl chains are known to be immunoreactive (Joyce & Van Kaer, 2003), and would therefore be expected to add to the immunoreactivity of the compounds of general formula I of the present invention.

DEFINITIONS

The term "alkyl" refers to the radical of saturated aliphatic groups, including straight-chain alkyl groups, branched-chain alkyl groups, cycloalkyl (alicyclic) groups, alkyl substituted cycloalkyl groups, and cycloalkyl substituted alkyl groups.

"Acyl" means an H--CO-- or alkyl-CO-- group wherein the alkyl group is as herein described.

Spacer or linker group E links together groups B and D of formula I of the present invention. By "spacer or linker group" is meant a group which covalently links a sugar moeity of group D to either a phosphate, phosphonate, carbamate, phosphothionate or sulfonate of group B. The linking group comprises alkyl chains which may be alicyclic, branched and/or further substituted with hydroxyl groups. The spacer/linker may have functionality which allows the attachment of one or more sugar chains. The spacer/linker may have a role of positioning the

95896-1

sugar moeities with respect to the group B phosphate, phosphonate, carbamate, phosphothionate, or sulfonate and the diacyl/dialkyl- or akyl-acyl- glyceryl unit of group A.

SYNTHESIS OF COMPOUNDS OF THE PRESENT INVENTION

5 **The synthesis of compound 7 (example 1a, Figure 1)**

Allyl α-D-mannopyranoside **1** was benzylated using benzyl bromide and sodium hydride in DMF (Lindhorst *et al.*, 2000) to give the mannoside **2** in 67% yield after purification by silica gel column chromatography. Ozonolysis of **2** and reductive workup with sodium borohydride gave, after purification by silica gel column chromatography, the alcohol **3** in 92% yield.

10 Treatment of **3** with H-phosphonate salt **4**, prepared as described by Crossman (Crossman *et al.*, 1997), and subsequent purification gave the triethylammonium salt **6** in a 57% yield. Hydrogenolytic debenzylation of **6** over 10% palladium on carbon in a solvent mixture comprising ethyl acetate, tetrahydrofuran, ethanol and water gave after purification over silica and lyophilization the target, compound **7** in 84% yield.

15

The synthesis of compound 15 (example 1b, Figure 2)

Benzylated allyl mannoside **2** was treated with a catalytic quantity of osmium tetraoxide and the re-oxidant, N-methyl morpholine-1-oxide to give the diol **8** as a 1 to 1 mixture of stereoisomers about the newly formed chiral centre (88% yield). Tritylation of the primary

20 hydroxyl group of **8**, subsequent benzoylation (benzoyl chloride and pyridine) and acid promoted detritylation gave benzoate **9** as a mixture of stereiosomers in 77% yield. Mannosylation of the primary hydroxyl group was achieved using phosphite **10** (Watanabe *et al.*, 1993; Watanabe *et al.*, 1994) promoted by N-iodosucinimide and trifluoromethane sulfonic acid in diethyl ether. The dimannoside **11** as a 4:1 mixture of alpha and beta anomers

25 at the new glycosidic linkage was obtained in 71% yield. Debenzoylation using sodium methoxide in methanol allowed isolation of the alpha-dimannoside **12** in 64% yield along with the alpha/beta dimannoside **13** (5%) and a mixture of **12** and **13** (12%).

Further examples satisfying the general structure, A-B-E-D could be synthesised by a skilled

30 worker via modification of the above synthetic procedures. Hydroboration of allyl glycoside **2** would give an alcohol (Lindhurst et al., 2000). Phosphorylation using the procedure

described for the synthesis of compound 7 would give a compound which comprises: A is a glyceride of formula Ia, where R_1 and R_2 are stearoyl, B is phosphate, E is $(CH_2)_n$, wherein n=3, D is a monosaccharide, namely a mannopyranose residue α-O-linked to a carbon atom of spacer E.

5

Further compounds may be synthesised where A is either a glyceride of formula Ia where R_1 and R_2 are different acyl groups or a combination of acyl and alkyl groups. Modification of diacylglyceryl groups is well documented (Hirth & Barner, 1982; Hirth *et al.*, 1983; Lindberg *et al.*, 2002).

10

Allyl glycosides of carbohydrates other than mannose are documented in the literature. Examples include allyl glycosides of disaccharides containing D-glucose residues (Koizumi *et al.*, 1991; Koto *et al.*, 1992).

15   Examples of compounds where the spacer is varied may be made from readily available polyols such as cyclohexanediols, erythritol and threoitol.

Compounds where the phosphate is replaced by other moieties may also be synthesised. For example, isocyanates derived from glycerol (A) are available (Green *et al.*, 1987). Reaction
20   of these with a D-E unit where D has a reactive hydroxyl group will give an A-B-E-D unit where B is $-NHCOO$ (ie carbamoyl).
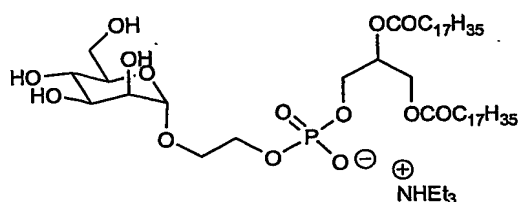
Particularly preferred synthetic molecules of the invention are:

25   **Compound 15**
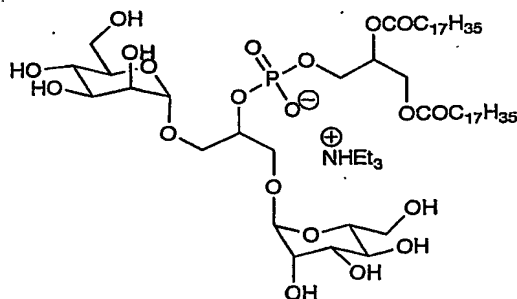
Compound 7

5   The synthesised molecules are each tested for biological activity in an animal model or in vitro model of disease as discussed below and suitably active compounds formulated into pharmaceutical compositions. The pharmaceutical compositions of the present invention may comprise, in addition to one or more synthetic molecules of the present invention, a pharmaceutically acceptable excipient, carrier, buffer, stabiliser or other material well known

10   in the art. Such materials should be non-toxic and should not interfere with the efficacy of the active ingredient. The precise nature of the carrier or other material will be dependent upon the desired nature of the pharmaceutical composition, and the route of administration e.g. oral, intravenous, cutaneous, subcutaneous, intradermal, nasal, pulmonary, intramuscular or intraperitoneal.

15

Pharmaceutical compositions for oral administration may be in tablet, lozenge, capsule, powder, granule or liquid form. A tablet or other solid oral dosage form will usually include a solid carrier such as gelatine, starch, mannitol, crystalline cellulose, or other inert materials generally used in pharmaceutical manufacture. Similarly, liquid pharmaceutical compositions

20   such as a syrup or emulsion, will generally include a liquid carrier such as water, petroleum, animal or vegetable oils, mineral oil or synthetic oil.

For intravenous, cutaneous, subcutaneous, intradermal or intraperitoneal injection, the active ingredient will be in the form of a parenterally acceptable aqueous solution which is pyrogen-

25   free and has suitable pH, isotonicity and stability.

For nasal or pulmonary administration, the active ingredients will be in the form of a fine powder or a solution or suspension suitable for inhalation. Alternatively, the active ingredients may be in a form suitable for direct application to the nasal mucosa such as an

30   ointment or cream, nasal spray, nasal drops or an aerosol.

A particularly preferred application of the biologically active compounds of the present invention is in the treatment of rhinitis. Rhinitis is an inflammatory disorder of the nasal passages. The symptoms of rhinitis typically consist of sneezing, rhinorrhea, nasal congestion and increased nasal secretion. Failure of treatment of rhinitis may lead to other disorders that include infection of the sinuses, ears and lower respiratory tract. To date, rhinitis is generally treated by oral medication comprising decongestants and antihistamines or mixtures thereof or by nasal administration of steroids, antihistamines or anti-cholinergics. However, such treatment is associated with various side effects including the sedating side effects of the antihistamines.

The present invention provides an oral pharmaceutical comprising at least one compound of the present invention together with a pharmaceutically acceptable carrier useful for the treatment of rhinitis.

Alternatively, the pharmaceutical composition may be formulated to deliver the active compound of the present invention directly to the mucosa of the nasal passages. The compounds of the present invention are particularly efficacious when delivered to the mucosal membranes due to their polar nature. Indeed, it is considered that the compounds of the present invention may form liposomes thereby providing an inherent delivery mechanism which may account, in part at least, for their potent biological activity (see results below). Preferred direct nasal mucosal delivery formulations include a nasal spray, nasal drops, cream or ointment.

Alternatively, rhinitis may be treated using a pharmaceutical composition of the present invention formulated for injection (either subcutaneous, cutaneous, intradermal, intramuscular or intraperitoneal injection).

For the treatment of asthma or other allergic respiratory disorders, the pharmaceutical compositions of the present invention may be formulated for respiratory administration to deliver the active ingredient to the airways of the patient to be treated. Generally, this will involve oral, intranasal or pulmonary delivery. Often, inhalation by the patient will provide

the motive force to deliver the active ingredient. However, respiratory administration can also involve delivery by propellant, including in the form of an aerosol generated using a jet or ultrasonic nebuliser as will be appreciated by a skilled worker.

5   In a further embodiment, the invention contemplates the use of one or more additional immuno-responsive compounds co-administered with the pharmaceutical composition of the present invention to give an additive or synergistic effect to the treatment regime. Such an immuno-responsive compound will generally be an immune response inducing substance. Examples of such substance include a natural lipo-aribomanan (LAM), a natural or synthetic

10   PIM, or mixtures thereof; glucocorticosteroids, such as prednisolone and methylprednisolone; nonsteroidal anti-inflammatory drugs (NSAIDs); as well as first and second generation anti-TNFα agents. Such substances may be administered either separately, sequentially or simultaneously with at least one compound of the present invention depending upon the condition to be treated as will be appreciated by a skilled worker.

15

Administration of the pharmaceutical composition of the invention is preferably in a "prophylactically effective amount" or a "therapeutically effective amount", this being sufficient to show benefit to the individual. The actual amount administered, and rate and time-course of administration, will depend on the nature and severity of what is being treated.

20   Prescription of treatment, e.g. decisions on dosage etc., is within the responsibility of general practitioners and other medical doctors, and typically takes account of the disorder to be treated, the condition of the individual patient, the site of delivery, the method of administration and other factors known to practitioners. Examples of the techniques and protocols mentioned above can be found in Remington's Pharmaceutical Sciences, 16th

25   edition, Oslo, A. (ed), 1980.

In addition, it is contemplated that the compounds of the present invention may be used as an adjuvant and may be formulated into adjuvant compositions by methods well known in the art.

30

The invention will now be described in greater detail by reference to specific Examples, which should not be construed as in any way limiting the scope of the invention.

95896-1

EXAMPLES

Reagents and Solvents

The following chemicals were purchased and used without further purification:

5   Laboratory grade solvents were used in this work. Dichloromethane was distilled from P_2O_5, THF from sodium wire (with benzophenone) and hexane was distilled using $CaCl_2$. All other reagents were purified according to the methods given in 'Purification of Laboratory Chemicals', 2nd ed. Perrin, D.D., Amarego, W. F. L. and Perrin, D. R., Peragamon Press Ltd, Oxford England (1981).

10

Salisyl chlorophosphite, (S)-(+)-1,3-dioxolane-4-methanol used for the synthesis of 1,2-*sn*-di-*O*-stearoyl glycerol, and D-(+)-mannose and methyl α-D-mannopyranoside used for the preparation of mannosyl donors, were purchased from the Aldrich Chemical company. 10% Palladium on carbon (Pd/C) was purchased from BDH.

15

Thin layer chromatography (TLC) was performed using aluminium-backed Merck Sorbent silica gel. Compounds were detected under an ultraviolet lamp and/or with a stain consisting of 5 % w/v dodecamolybdophosphic acid in ethanol, followed by development with a heat gun. Column chromatography was performed using silica gel (Sorbasil, particle size 32-63

20   μm), which was packed by the slurry method.

Instrumentation:

Nuclear Magnetic Resonance (NMR) Spectroscopy

^{1}H NMR spectra were recorded at either 300 MHz on a Varian Unity Inova 300 MHz

25   spectrometer or at 500 MHz on a Varian Unity Inova 500 MHz spectrometer. All spectra were recorded in the stated solvent at 25 °C in 5 mm NMR tubes. Chemical shifts are reported relative to $CHCl_3$ at 7.26 ppm using the δ scale. Chemical shifts have an uncertainty of ± 0.01 ppm. Coupling constants (*J*) have been rounded to the nearest 0.5 Hz. Resonances were assigned as follows: chemical shift (number of protons, multiplicity, coupling constant(s),

assigned proton(s)). Multiplicity abbreviations are reported by the conventions: s (singlet), d (doublet), dd (doublet of doublets), t (triplet), m (multiplet).

^{13}C NMR spectra were recorded at either 75 MHz on a Varian Unity Inova 300 MHz spectrometer or at 125 MHz on a Varian Unity Inova 500 MHz spectrometer. Chemical shifts of carbon nuclei are reported relative to $CDCl_3$ at 70.08 ppm.

^{31}P NMR spectra were recorded at either 131 MHz on a Varian Unity Inova 300 MHz spectrometer or at 202 MHz on a Varian Unity Inova 500 MHz spectrometer. Chemical shifts of phosphorous nuclei are reported relative to 80% H_3PO_4 as an external standard at 0.0 ppm.

Infrared (IR) Spectroscopy

Infrared spectra were recorded on a Perkins Elmer 1600 series FTIR spectrophotometer. Samples were examined as thin films between two NaCl plates.

Microanalyses

Microanalyses were carried out by the Campbell Microanalytical Laboratory, University of Otago, Dunedin, New Zealand.

Mass Spectroscopy (MS).

Low resolution mass spectra were run on a Shimadzu QP8000 alpha with APCI or ESI probes. ESI spectra were run using 9:1 acetonitrile/water mobile phase, CDL temperature of 250 °C and a cone voltage of 50 eV. APCI spectra were run using 1:1 methanol/water mobile phase, CDL temperature of 250 °C, APCI temperature of 400 °C and a cone voltage of 50 eV. High resolution mass spectra were run in the ESI/ mode on MicroMass LCT coupled to a Waters 2790 LC with a 996 PDA, source 80 °C probe temperature as required for solvent scanning 2500-100AMU at 1/sec with a Cole Palmer syringe pump for direct infusion work at the Department of Chemistry, University of Canterbury, New Zealand.

Polarimetry

Optical rotations were recorded on a Jasco DIP-100 digital polarimeter using a 1 dm cell.

Chromatography

5   Thin layer chromatography (TLC) was performed using aluminium-backed Merck Sorbent silica gel. Compounds were detected under an ultraviolet lamp and/or with a stain consisting of 5 % w/v dodecamolybdophosphic acid in ethanol, followed by development with a heat gun.

10   Column chromatography was performed using silica gel (Sorbasil, particle size 32-63 μm), which was packed by the slurry method.

EXAMPLE 1:   **Synthesis of Compounds Corresponding to General Formula (I)**

15

EXAMPLE 1(a):   **Synthesis of triethylammonium 1-*O*-(1,2-distearoyl-*sn*-glycero-3-phosphoryl)-2-*O*-(α-D-mannopyranosyl)-1,2-ethanediol 7 (Compound 7)**

The total synthesis of Compound 7 is represented schematically in Figure 1.

20

Synthesis of Allyl 2,3,4,6-tetra-*O*-benzyl-α-D-mannopyranoside 2 (Lindhurst *et al*; 2000)

2
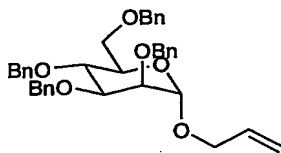
25   Allyl α-D-mannopyranoside 1(Gigg *et al.*, 1985; RajanBabu *et al.*, 1989) (1.50 g, 2.78 mmol) was suspended in benzyl chloride (40 mL) and sodium hydride (60%, 2.1 g, 52.5 mmol) was carefully added. The suspension was stirred at 125 °C for 6 h under nitrogen. The mixture was filtered and excess benzyl chloride was distilled off under reduced pressure. The residue was

dissolved in dichloromethane (100 mL), washed with water (300 mL) and dried (MgSO$_4$). After removal of the solvent the residue was purified over silica (hexane/ether 4:1 as eluent) to give the title compound **2** (3.2 g, 67%) as pale yellow syrup; $[\alpha]_D^{21.5}$ +25.0 (*c* 1.5, CH$_2$Cl$_2$); ^{1}H NMR (500 MHz, CDCl$_3$) δ 7.40-7.25 and 7.18-7.13 (m, 20H, Ar-H), 5.88-5.79

5   (m, 1H, H-2′), 5.21 (ddd, 1H, *J* 17, 3, 1.5 Hz, H-3a′), 5.18 (ddd, 1H, *J* 10.5, 3, 1 Hz, H-3b′), 4.92 (d, 1H, *J* 2 Hz, H-1), 4.88 (d, 1H, *J* 10.5 Hz, PhCH$_2$), 4.75 (d, 1H, *J* 12.5 Hz, PhCH$_2$), 4.17 (d, 1H, *J* 12.0 Hz, PhCH$_2$), 4.67 (d, 1H, *J* 12.0 Hz, PhCH$_2$), 4.63 (s, 2H, PhCH$_2$), 4.55 (d, 1H, *J* 12 Hz, PhCH$_2$), 4.50 (d, 1H, *J* 10.5 Hz, PhCH$_2$), 4.16 (ddt, 1H, *J* 13.5, 4.5, 1.5 Hz, H-1′a), 4.02-3.92 (m, 3H), 3.83-3.71 (m, 4H); ^{13}C NMR (125 MHz, CDCl$_3$) δ 138.6, 138.5,

10   138.5, 138.4 (C$_q$-Ar), 133.9 (C2′), 128.4, 128.4, 128.3, 128.1, 127.9, 127.8, 127.7, 127.64, 127.62, 127.59, 127.5 (CH-Ar), 117.3 (C3′), 97.2 (C1, J_{C1-H1} 169 Hz), 80.3 (C1′), 75.2, 75.0, 74.7, 73.4, 72.6, 72.2, 72.0, 69.3, 67.9; ESI-MS (+ve ion) *m/z*(%) 604[MNa+1]$^+$ (71), 603[MNa]$^+$ (100).

15   · **Synthesis of 2-Hydroxyethyl 2,3,4,6-tetra-*O*-benzyl-α-D-mannopyranoside 3**
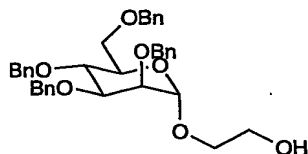

3

Ozone gas was bubbled through a solution of allyl 2,3,4,6-tetra-*O*-benzyl-α-D-mannopyranoside **2** (1.0 g, 1.72 mmol) in methanol (100 mL) at −78 °C until a slight blue colour persisted (2-3 minutes). The reaction was warmed to room temperature, sodium

20   borohydride (460 mg, 12.1 mmol) was added and the mixture was stirred for one hour. The solvent was distilled off and the residue was treated with 2M HCl (30 mL). The compound was then extracted in dichloromethane, dried over magnesium sulfate and solvent removed *in vacuo*. The residue was purified over silica (hexane/ether, 2:3 as eluent) to give the title

25   compound **3** (916 mg, 92%) as colourless syrup; Rf 0.4 (ether/hexane, 3:2); $[\alpha]_D^{21.5}$ +14.7 (*c* 2.85, CH$_2$Cl$_2$); (Found: C, 73.84; H, 6.97. C$_{36}$H$_{40}$O$_7$ requires C, 73.95; H, 6.97); ^{1}H NMR (500 MHz, CDCl$_3$) δ 7.37-7.26 and 7.16-7.14 (m, 20 Hs, Ar-H), 4.92 (d, 1H, *J* 2 Hz, H-1), 4.90 (d, 1H, *J* 11 Hz, PhCH$_2$), 4.78 (d, 1H, *J* 11 Hz, PhCH$_2$), 4.74 (d, 1H, *J* 11 Hz, PhCH$_2$), 4.66 (s, 2H, PhCH$_2$), 4.64 (d, 1H, *J* 11.5 Hz, PhCH$_2$), 4.57 (d, 1H, *J* 11.5 Hz, PhCH$_2$), 4.52 (d,

30   1H, *J* 11 Hz, PhCH$_2$), 3.96-3.84 (m, 5H), 3.75-3.63 (m, 5H); ^{13}C NMR (125 MHz, CDCl$_3$) δ

138.6, 138.4, 138.4, 138.2 (C_q-Ar), 128.5, 128.46, 128.1, 127.9, 127.8, 127.75, 127.7 (CH-Ar), 98.9 (C-1), 80.1 (CH), 75.2 (CH CH_2), 75.0 (CH), 73.5 (CH_2), 72.9 (CH_2), 72.4 (CH_2), 72.2 (CH), 70.9 (CH_2), 69.4 (CH_2), 62.1 (CH_2); ESI-MS (+ve ion) m/z(%) 607[MNa]$^+$ (100).

5 **Synthesis of _1,2_-Distearoyl-_sn_-glycero-3-H-phosphonate triethylammonium salt 4 (Crossman _et al.,_ 1997)**


4   **5**
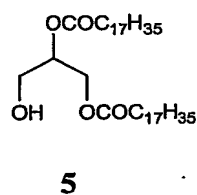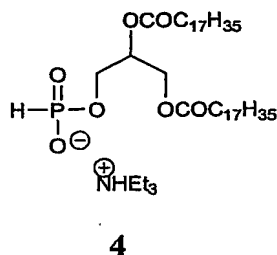
10

1,2-Distearoyl-_sn_-glycerol 5 (150 mg, 0.24 mmol) prepared by the method of Hirth (Hirth & Barner, 1982) was dried by evaporation from pyridine and was dissolved in pyridine/THF (2 mL, 1:4). The solution was then added dropwise to the stirred solution of salicyl chlorophosphite (80mg, 0.396 mmol) in THF (2mL). The reaction was stirred at room temperature for 15 minutes and 1M aqueous triethylammonium bromide (TEAB) solution (10 mL) was added followed by the addition of chloroform (10 mL). The organic layer was washed with water (25 mL), 1M TEAB (20 mL) and dried over magnesium sulfate. Removal of the solvent gave salt **4** which was used without further purification in the following reaction.

20

Synthesis of Triethylammonium 1-_O_-(1,2-disteroyl-_sn_-glycero-3-phoshoryl)-2-_O_-(2,3,4,6-tetra-_O_-benzyl-α-D-mannopyranosyl)-1,2-ethanediol 6

6
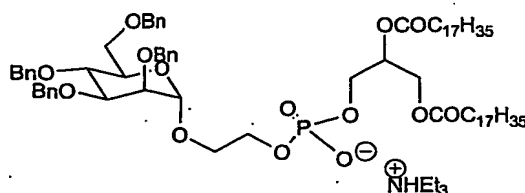
25

Salt **4** and mannopyranoside **3** (102 mg, 0.18 mmol) were dried by co-evaporation with pyridine (2 × 20 mL). The mixture was dissolved in pyridine (40 mL), pivaloyl chloride (196µL, 1.59 mmol) was added and the reaction mixture was stirred for 1 h at room temperature. A solution of iodine (132 mg, 0.52 mmol) in a 9:1 mixture of pyridine/water (10 mL) was added and stirring was continued for 45 min. The reaction mixture was diluted with dichloromethane (50 mL), washed with 10% sodium thiosulfate solution (20 mL), with 1M TEAB (2 × 20 mL) and water (100 mL). The organic layer was dried over magnesium sulfate and the solvent was removed. The residue was purified over silica (dichloromethane/methanol/TEA 97:2:1 as eluent) to give the title compound **6** (176 mg, 57%) as clear glass; R_f 0.45 (methanol/ether/dicholoromethane, 1:2:7); $[\alpha]_D^{21.5}$ +7.3 (c 0.3, CH_2Cl_2); ^{1}H NMR (500 MHz, $CDCl_3$) δ 7.38-7.23 and 7.15-7.14 (m, 20 H, Ar-H), 5.24-5.18 (m, 1H, H-2″), 4.96 (d, 1H, J 2.5 Hz, H-1), 4.86 (d, 1H, J 11 Hz, $PhCH_2$), 4.73 (s, 2H, $PhCH_2$), 4.72 (d, 1H, J 12.5 Hz, $PhCH_2$), 4.67 (d, 1H, J 12 Hz, $PhCH_2$), 4.60 (d, 2H, J 3.0 Hz, $PhCH_2$), 4.51 (t, 2H, J 14.5 Hz), 4.40-4.36 (m, 1H), 4.18-4.14 (m, 1H), 4.03-3.95 (m, 5H), 3.92-3.87 (m, 1H), 3.87-3.86 (m, 1H), 3.82-3.70 (m, 3H), 3.64-3.62 (m, 1H), 2.84 (q, 6H, J 7.0 Hz, 3 × NCH_2), 2.25 (t, 4H, J 8.0 Hz, 2 × $COCH_2$), 1.58-1.54br (m, 4H), 1.30-1.11br (m, 65H), 0.88 (t, 6H, J 10.5 Hz, 2 × CH_3); ^{13}C NMR (125 MHz, $CDCl_3$) δ 173.7, 173.3, 138.92, 138.87, 138.74, 138.71 (C_q-Ar), 128.55, 128.52, 128.48, 128.27, 128.04, 127.99, 127.75, 127.72, 127.68 (CH-Ar), 98.3 (C-1), 80.6, 75.3, 75.1, 74.9, 73.6, 72.8, 72.2, 72.1, 70.7, 69.5, 67.4, 64.5, 63.0, 45.9, 34.6, 34.4, 32.2, 30.0, 29.9, 29.8, 29.62, 29.60, 29.58, 29.41, 29.39, 25.2, 25.1, 22.9, 14.4, 9.7; ^{31}P NMR (202 MHz, $CDCl_3$) δ 0.607 ppm; ESI-MS(-ve ion) m/z(%) 1270(100), 1269(87); HRMS-ESI(-ve) (Found: m/z 1269.7978. $C_{75}H_{114}O_{14}P^-$ requires m/z 1269.7946).

Synthesis of Triethylammonium 1-*O*-(1,2-distearoyl-*sn*-glycero-3-phosphoryl)-2-*O*-(α-D-mannopyranosyl)-1,2-ethanediol 7 (Compound 7)

7
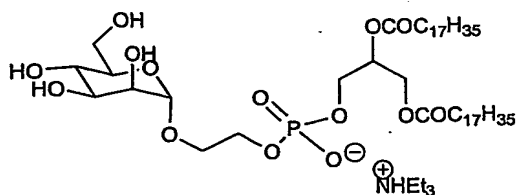
Phosphate **6** (60 mg, 0.047 mmol) was dissolved in a 2:1:1:1 mixture of EtOAc/THF/EtOH/H$_2$O (20 mL). 10% Pd/C (120 mg, BDH) was added and the reaction was stirred under an atmosphere of hydrogen for 18 h. After filtration through Celite, the filter pad was washed with THF (5 mL) and dichloromethane (2 × 5mL) and the solvent was removed. Water was removed by azetropic distillation with toluene (5 × 3mL). The residue was purified over silica (dicholoromethane/methanol/TEA 94:5:1 as eluent) to give, after lyophilization from methanol and water, the title compound **7** (36 mg, 84%) as a white solid; Rf 0.2 (dicholoromethane:methanol/TEA 9:1:1); [$\propto$]$_D^{21.5}$ +10 (c 0.9, CH$_2$Cl$_2$); ^{1}H NMR (500 MHz, CDCl$_3$) δ 5.22*br* (m, 1H, H-C2″), 4.92*br* (1H, H-1), 4.38*br* (d, 1H, J 2.5 Hz, H-3″), 4.16 (dd, J 2.5, 1.15 Hz, H-1″ and 3″), 4.08-3.58 (m, 12H), 3.73*br* (s, 6H, 3 × NCH$_2$), 2.34-2.26 (m, 4H, 2 × COCH$_2$), 1.60-1.50*br* (m, 4H), 1.19-1.29*br* (m, 65 H), 0.90 (t, 6H, J 10.5 Hz, 2 × CH$_3$); ^{13}C NMR (125 MHz, CDCl$_3$) δ 173.5, 173.2, 100.2 (C-1), 72.7, 71.7, 70.8, 70.5, 70.4, 67.9, 67.0, 65.0, 63.6, 62.7, 62.1, 45.8, 34.4, 34.2, 32.0, 29.8, 2′ ̇ ̦, 29.6, 29.43, 29.41, 29.24, 29.23, 25.0, 24.95, 22.8, 14.2, 8.9; ^{31}P NMR (202 MHz, CDCl$_3$) δ -0.229 ppm; ESI-MS(–ve ion) m/z(%) 910(100), 909(60); HRMS, ESI (-ve) (Found: m/z 909.6078. C$_{47}$H$_{90}$O$_{14}$P$^-$ requires m/z 909.6068).

EXAMPLE 1(b): **Synthesis of triethylammonium (1,2-disteroyl-*sn*-glycero-3-phoshoryl)-2-[1,3-*bis*-(α-D-mannopyranoside)]glycerol 15 (Compound 15)**

The total synthesis of compound 15 is represented schematically in Figure 2.

<u>Synthesis of 1-*O*-(2,3,4,6-Tetra-*O*-benzyl-α-mannopyranosyl)glycerol **8**</u>

8
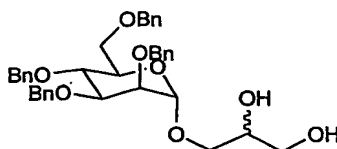
Allyl 2,3,4,6-tetra-*O*-benzyl-α-D-mannopyranoside **2** (1.00 g, 1.72 mmol) and *N*-methylmorpholine-1-oxide (302 mg, 2.58 mmol) were dissolved in a mixture of acetone/water 9:1 (40 mL) and a 1% aqueous osmium tetraoxide solution (1.5 mL) was added. The reaction mixture was stirred overnight at room temperature, then poured into 10% sodium thiosulfate

5 solution (20 mL) and extracted with dicholoromethane (40 mL). The organic layer was washed with water and dried over magnesium sulfate. The solvent was removed and the residue was purified over silica (hexane/ether 1:2 as eluent) to give the title compound **8** (927 mg, 88%, 1:1 mixture of epimers) as a colourless syrup; R*f* 0.4 (hexanes/ethylacetate, 1:2); (Found: C, 71.69; H, 6.92. $C_{37}H_{42}O_8.0.5H_2O$ requires C, 71.25; H, 6.95; O, 21.80); v_{max}/cm^{-1}

10 3439 (OH); ^{1}H NMR (500 MHz, CDCl$_3$) δ 7.36-7.24 and 7.21-7.18 (m, 20H, Ar-H), 4.87-4.71 (d, 1H, *J* 12.5 Hz, PhCH$_2$), 4.64-4.59 (m, 1H), 4.55 and 4.53 (2 x d, 1H, *J* 12.5 Hz, PhCH$_2$), 4.49 (dd, 1H, *J* 11 and 4.5 Hz, PhCH$_2$), 3.94-3.46 (m, 11H); ^{13}C NMR (125 MHz, CDCl$_3$) δ 138.4, 138.3, 138.24, 138.23, 138.1, 138.0 (C$_q$-Ar), 128.45, 128.43, 128.40, 128.10, 128.06, 127.99, 127.95, 127.90, 127.76, 127.72, 127.70, 99.1 (C-1), 79.9, 75.12, 75.10, 75.04,

15 75.02, 74.9, 73.6, 73.5, 72.9, 72.85, 72.42, 72.37, 72.3, 72.2, 70.9, 70.8, 70.6, 69.9, 69.5, 69.4, 63.6, 63.5; ESI-MS (+ve ion) *m/z* (%) 638[MNa+1]$^+$ (24), 91 (100).

Synthesis of 2-*O*-Benzoyl-3-*O*-(2,3,4,6-tetra-*O*-benzyl-α-D-mannopyranosyl)glycerol **9**
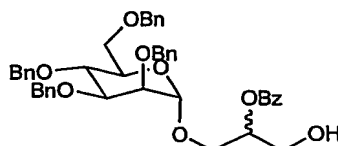
9

5   Glycerol **8** (900 mg, 1.47 mmol) and trityl chloride (490 mg, 1.76 mmol) were dissolved in dry pyridine (60 mL) and heated at 100 °C. The disappearance of **8** was monitored by TLC and after 90 min the reaction was cooled to 0 °C and a solution of benzoyl chloride (1.0 g, 7.0 mmol) in dry dichloromethane (10 mL) was added. The reaction was warmed to room temperature and stirring was continued for 2 h. The solvent was removed and the residue was

10   dissolved in chloroform (50 mL), washed with 2M HCl (2 × 20 mL), saturated sodium bicarbonate solution (2 × 20 mL), water (25 mL) and dried over magnesium sulfate. The solvent was removed, the residue was dissolved in mixture of dicloromethane and methanol (7:3, 50 mL) and *p*-TSA (75 mg) was added. The reaction was stirred at room temperature overnight and solvent was removed. The residue was purified over silica [hexane/ether, 8:2 to

15   7:3 as eluent] to afford the title compound **9** (816 mg, 77%) as a pale syrup; R*f* 0.4 (ether/hexane, 2:1); ^{1}H NMR (500 MHz, CDCl$_3$) δ 8.05*br* (d, 3H, *J* 7.5 Hz), 7.51 (t, 2H, *J* .5 Hz), 7.41-7.12 and 7.18-7.10 (m, 20 Hs, Ar-H), 5.30-5.20 (m, 1H, H-2'), 4.94 and 4.89 (2 x d, each 1H, *J* 2Hz, H-1), 4.85 and 4.83 (2 x d, each 1H, *J* 10.5 Hz, PhCH$_2$), 4.77-4.46 (m, 7H), 4.05-3.62 (m, 10H); ^{13}C NMR (125 MHz, CDCl$_3$) δ 166.1, 166.0 (CO), 138.3, 138.2, 138.1,

20   138.05 (C$_q$-Ar), 133.0, 132.96, 129.8, 129.7, 129.6, 129.5, 128.3, 128.2, 128.2, 128.13, 128.10, 128.07, 127.83, 127.80, 127.74, 127.65, 127.62, 127.59, 127.56, 127.41, 127.38, 127.36, 127.32 (CH-Ar), 98.0 and 97.5 (C-1), 79.7, 79.6, 74.8, 74.7, 74.6, 73.8, 73.4, 73.2, 73.1, 72.4, 72.0, 71.9, 69.0, 68.9, 65.6, 65.3, 65.25, 61.2, 57.9; ν$_{max}$/cm^{-1}(CHCl$_3$) 1716; ESI-MS (+ve) *m/z* 742[MNa]$^+$ (100); HRMS-ESI (+ve) (Found: *m/z* 719.3220 (MH$^+$)). C$_{44}$H$_{47}$O$_9$

25   requires *m/z* 719.3220.

Synthesis of (2,3,4,6-Tetra-*O*-benzyl-D-mannopyranosyl)dimethylphosphite **10** (Watanabe *et al.*, 1993; Watanabe *et al.*, 1994)
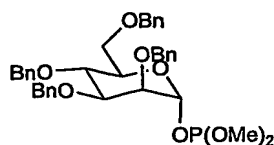

10

5

A mixture of 2,3,4,6-tetra-*O*-benzyl-D-mannose (Koto *et al.*, 1976) (940 mg, 1.74 mmol), dimethoxy-*N,N*-diisopropylphosphoromidate (437 mg, 2.27 mmol) and 4,5-dichloroimidazole (355 mg, 2.61 mmol) in dry dichloromethane (25 mL), under nitrogen, was stirred at room temperature for 105 min. The mixture was poured into water (100 mL) and extracted with

10   dichloromethane. The organic layer was washed with water, dried over magnesium sulfate and the solvent was removed. The residue consisting mainly of phosphite **10** was used without further purification.

Synthesis of 2-*O*-Benzoyl-1,3-*bis-O*-(2,3,4,6-tetra-*O*-benzyl-α-D-mannopyranosyl)glycerol **11**
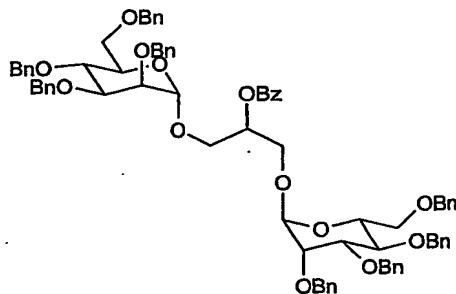

11

15

Powdered 4Å molecular sieves (50 mg) were added to a solution of glycerol **9** (730 mg, 1.02 mmol), phosphite **10** (771 mg, 1.22 mmol) and N-iodosuccinimide (NIS) (275 mg, 1.22 mmol) in dry ether (20 mL) under an atmosphere of nitrogen. The reaction was stirred at room temperature for 15 minutes, trifluoromethanesulfonic acid (110 μL, 1.24 mmol) was added

20   and the stirring was continued for 2 h. The reaction was diluted with more ether (50 mL) and the organic layer was washed with 10% sodium thiosulfate solution (30 mL), water (2 × 20 mL) and dried over magnesium sulfate. The solvent was removed and the residue was purified over silica [hexane/ether 9:1 as eluent] to give **11** (900 mg, 71 %, α to β 4:1) as a colourless syrup; R*f* 0.6 (hexane/ether, 2:3); v_{max}cm^{-1}/(CHCl$_3$) 1716.7; ^{1}H NMR (500 MHz, CDCl$_3$) δ

25   *inter alia* 8.00 (d, *J* 7.7, 2H), 7.55 (t, *J* 7.7 Hz, 1H), 7.38-7.08 (m, 40H, Ar-H), 5.53-5.41 and

5.38-4.43 (m, together 1H, H-2'), 4.96 (d, 1H, J 2 Hz, H-1), 4.91 (d, 1H, J 2 Hz, H-1), 4.86 (d, 2H, J 11 Hz, PhCH$_2$), 4.82 (d, 2H, J 11 Hz, PhCH$_2$), 4.72 (d, 2H, J 12.5 Hz, PhCH$_2$), 4.69 (d, 2H, J 12.5 Hz, PhCH$_2$), 4.65-4.55 (m, 4H), 4.52-4.42 (m, 4H), 4.13-3.50 (m, 16H); ^{13}C NMR (125 MHz, CDCl$_3$) δ *inter alia* 165.9 (CO), 138.6, 138.5, 138.5, 138.43, 138.42, 138.35 (C$_q$-Ar), 133.2 (C$_q$-benzoyl), 130.0, 129.8, 128.5, 128.41, 128.35, 128.33, 128.29, 128.03, 127.92, 127.87, 127.81, 127.78, 127.76, 127.73, 127.63, 127.60, 127.52, 98.4 and 97.7 (2 x C-1), 80.0, 79.9, 75.1, 75.0, 74.9, 74.8, 74.7, 73.42, 73.38, 72.68, 72.65, 72.4, 72.3, 72.2, 71.4, 69.2, 69.1, 65.9, 65.5; ESI-MS (+ve) *m/z* (%) 1265 [MNa+1]$^+$(18), 1264 (MNa$^+$,47), 91 (100); HRMS-ESI (+ve) (Found: *m/z* 1241.5595 (MH$^+$)). C$_{78}$H$_{81}$O$_{14}$ requires *m/z* 1241.5626.

Synthesis of 1, 3-*Bis-O*-(2,3,4,6-tetra-*O*-benzyl-α-D-mannopyranoyl)glycerol **12**


12

13
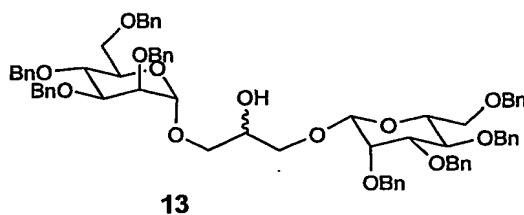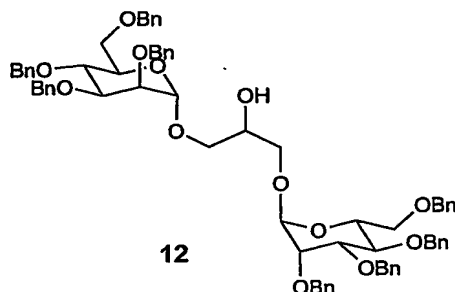
Dimannoside **11** (800 mg, 0.65 mmol) was dissolved in 1M sodium methoxide in methanol (80 mL) and the reaction was stirred overnight at room temperature. The solvent was removed and the residue was dissolved in dichloromethane, washed with 2M HCl (2 x 50 mL) and water (50 mL), and dried over magnesium sulfate. Removal of the solvent gave a residue (α to β 4:1, 615 mg) which was purified over silica [hexane/ether 75:25 to 65:35 gradient elution] to afford the title compound **12** (470 mg, 64%), a mixture of **12** and **13** (90 mg, 12%), and **13** (40 mg, 5%).

Data for **12**: R*f* 0.55 (hexane/ether, 2:3); $[\propto]_D^{21.5}$ +24 (*c* 1.05, CH$_2$Cl$_2$); ν_{max}cm^{-1}/(CHCl$_3$) 3018*br* (OH); ^{1}H NMR (500 MHz, CDCl$_3$) δ 7.35-7.20 and 7.15-7.12 (m, 40H, Ar-H), 4.85-4.82 (m, 4H), 4.71 (d, 2H, *J* 12.5 Hz, PhCH$_2$), 4.67 (d, 2H, *J* 12.5 Hz, PhCH$_2$), 4.62-4.55 (m, 6H), 4.49-4.45 (m, 4H), 3.96 (d, 1H, *J* 10 Hz), 3.91(d, 1H, *J* 10 Hz), 3.77-3.65 (m, 8H), 3.60-3.48 (m, 6H), 3.44-3.39 (m, 1H); ^{13}C NMR (125 MHz, CDCl$_3$) δ 138.52, 138.45, 138.43, 138.34, 138.29 (C$_q$-Ar), 128.42, 128.41, 128.40, 128.38, 128.36, 128.05, 127.84, 127.83, 127.72, 127.69, 127.67, 127.64, 127.59, 127.56 (CH-Ar), 98.55 and 98.58 (both C-1, ^{1}*J*C-H 170 Hz), 80.1, 80.0, 75.1, 75.08, 74.91, 74.77, 73.44, 73.42, 72.76, 72.37, 72.33, 72.26, 72.24, 70.0, 69.6, 69.4, 69.24, 69.21; ESI-MS (+ve) 1177 [MK+1]$^+$ (59), 1176 [MK]$^+$ (78), 1161 [MNa+1]$^+$ (97), 1160 [MNa]$^+$ (100); HRMS-ESI (+ve) (Found: *m/z* 1159.5200 MNa$^+$). C$_{71}$H$_{76}$O$_{13}$Na$^+$ requires *m/z* 1159.5184.

Data for **13**: R*f* 0.5 (hexane/ether, 2:3); ν_{max}cm^{-1}/(CHCl$_3$) 3012*br* (OH); ^{1}H NMR (500 MHz, CDCl$_3$) δ 7.34-7.16 m, 40H, Ar-H), 5.04 (d, 1H, *J* 2Hz, H-1α), 4.92-4.83 (m, 2H), 4.76-4.44 (m, 15H), 4.00-3.64 (m, 16H), 3.58-3.42 (m, 1H); ^{13}C NMR (125 MHz, CDCl$_3$) δ 138.53, 138.50, 138.42, 138.36, 138.30, 138.26, 138.23 (C$_q$-Ar), 128.40, 128.37, 128.22, 128.18, 128.14, 128.08, 128.06, 127.97, 127.92, 127.84, 127.76, 127.69, 127.66, 127.59, 127.55, 98.6 and 97.7 (both C-1), 80.14, 79.95, 75.17, 75.11, 74.96, 74.92, 74.88, 73.46, 72.78, 72.71, 72.40, 72.38, 72.36, 72.23, 69.29, 67.03, 61.69; ESI-MS (+ve) 1177 [MK+1]$^+$ (35), 1176 [MK]$^+$ (100), 1160 [MNa]$^+$ (89); (Found: C, 74.87; H, 6.86; C$_{71}$H$_{76}$O$_{13}$ requires C, 74.98; H, 6.74).

<u>Synthesis of Triethylammonium 2-*O*-(1,2-distearoyl-*sn*-glycero-3-phosphoryl)-1,3-*bis-O*-(2,3,4,6-tetra-*O*-benzyl-α-D-mannopyranosyl)glycerol **14**</u>

14
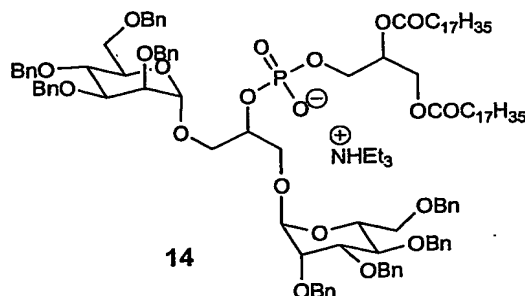
A mixture of salt 4, prepared from **5** (200 mg, 0.32 mmol), salicyl chlorophosphite (107 mg, 0.53 mmol) using the procedure previously described, and glycerol **12** (200 mg, 0.176 mmol) was dried by co-evaporation with pyridine (2 × 20 mL) and was then dissolved in pyridine (8 mL). Pivaloyl chloride (200 μL, 1.62 mmol) was added and the resulting solution was stirred

5   for 1 h at room temperature. After this time a solution of iodine (120 mg, 0.47 mmol) in a 9:1 mixture of pyridine/water (30 mL) was added and stirring was continued for 45 min. The reaction mixture was diluted with dichloromethane (50 mL) and stirred for 15 min, washed with 10% sodium thiosulfate solution (20 mL), and with 1M TEAB (2 × 20 mL) and water (100mL). The organic layer was dried over magnesium sulfate and the solvent was removed.

10 The residue was purified over silica (dichloromethane/methanol/TEA 98:1:1 as eluent) to give the title compound **14** (252 mg, 76 %) as clear glass; Rf 0.45 (dichloromethane/methanol/TEA, 5:1:0.1); $[\propto]_D^{22.1}$ +11.7 (c 1.2, CH$_2$Cl$_2$). ^{1}H NMR (500 MHz, CDCl$_3$) δ 7.38-7.11 (m, 40H, Ar-H), 5.22-5.18 (m, 1H, H-1"), 5.03 (d, 1H, J 6 Hz, H-1), 4.96br (s, 1H, H-1), 4.86-4.80 (m, 2H, PhCH$_2$), 4.74-4.25 (m, 15H, PhCH$_2$), 4.15-4.10 (m,

15 1H), 4.06-3.92 (m, 5H), 3.90-3.62 (m, 14H), 2.90-2.80br (m, 6H, 3 × NCH$_2$), 2.21 (t, 4H, J 7.5 Hz, 2 × COCH$_2$), 1.58-1.44br (m, 4H, 2 × COCH$_2$CH$_2$), 1.32-1.20 (m, 48H), 1.15 (t, 9H, J 7.5 Hz, 3 × NCH$_2$CH$_3$), 0.88 (t, 6H, J 7.5 Hz, 2 × CH$_3$); ^{13}C NMR (125 MHz, CDCl$_3$) δ 173.40, 172.99 (CO), 138.76, 138.74, 138.66, 138.58 (C$_q$-Ar), 128.35, 128.27, 128.24, 128.22, 128.04, 128.03, 127.80, 127.74, 127.71, 127.67, 127.65, 127.60, 127.58, 127.45,

20 127.38 (CH-Ar), 98.29 (C-1'), 80.47, 80.39, 75.08, 75.04, 74.80, 74.75, 73.37, 73.33, 72.83, 72.54, 72.08, 72.02, 70.52, 69.32, 69.21, 66.97, 63.58, 62.95, 45.43, 34.29, 34.11, 31.96, 29.75, 29.70, 29.57, 29.40, 29.37, 29.20, 24.96, 24.89, 22.73, 14.16, 8.71; ^{31}P NMR (121 MHz, CDCl$_3$) δ 0.15 ppm; ESI-MS (–ve) m/z(%) 1824(38), 1823(90), 1822(100). –? HRMS-ESI (-ve) (Found: m/z 1823.0492); C$_{116}$H$_{166}$NO$_{20}$P$^-$ requires m/z 1823.0526.

25

Synthesis of Triethylammonium (1,2-disteroyl-*sn*-glycero-3-phoshoryl)-2-[1,3-*bis*-(α-D-mannopyranoside)]glycerol **15** (Compound 15)

15
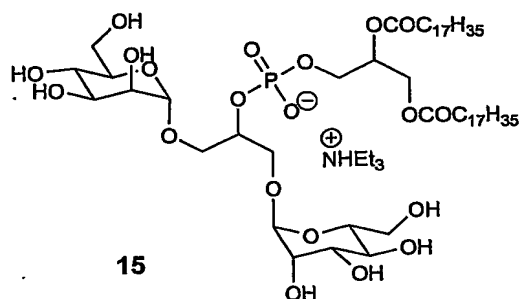
5 Phosphate **14** (100 mg, 0.055 mmol) was dissolved in a 2:1:1:1 mixture of EtOAc/THF/EtOH/H$_2$O (30 mL). 10% Pd/C (200 mg) was added and the reaction was stirred under the atmosphere of hydrogen for 18 h. The mixture was filtered through Celite, the filter pad was washed with THF (5 mL), methanol (5 mL) and dichloromethane (2 x 5 mL), and the solvent from the combined filtrates was removed *in vacuo*. The water was removed by

10 azeotropic distillation with toluene (5 × 4mL). The residue was purified by silica gel preparative plate chromatography (dichloromethane/methanol/TEA 94:5:1 as eluent). The baseline region of the plate was cut and the silica washed with warm methanol (20 mL) and dichloromethane (20 mL). The solvent was removed to give a residue which was lypophilised from a methanol and water mixture to give the title compound **15** (42 mg, 69%) as a white

15 solid; [∝]$_D^{22.1}$ +31 (c 0.3, CH$_2$Cl$_2$); ^{1}H NMR (500 MHz, CDCl$_3$/CD$_3$OD/D$_2$O 35:20:3) δ 5.10-4.90 (m, 1H, H-2"), 4.59 (dd, 1H, *J* 4 and 2 Hz, H-1), 4.56*br* (s, 1H, H-1), 4.82-3.76 (m, 9H), 3.74-3.69 (m, 2 H), 3.63-3.57 (m, 4H), 3.57-3.32 (m, 6H), 2.87 (q, 6H, *J* 7.5 Hz, 3 × NCH$_2$), 2.10-2.04 (m, 4H, 2 × COCH$_2$), 1.42-1.25*br* (m, 4H, 2 × COCH$_2$C*H$_2$*), 1.15-0.95*br* (m, 65H), 0.63 (t, 6H, *J* 7 Hz, 2 × CH$_3$); ^{13}C NMR (125 MHz, CDCl$_3$/CD$_3$OD/D$_2$O 35:20:3) δ

20 173.9, 173.5 (CO), 100.0, 99.9, 90.5, 72.7, 72.6, 70.8, 70.1, 67.03, 66.98, 66.4, 66.0, 63.1, 62.5, 61.1, 58.6, 48.6, 48.4, 48.3, 48.1, 47.9, 47.8, 47.6, 46.0, 33.9, 33.7, 31.5, 29.3, 29.24, 29.19, 29.15, 29.00, 28.95, 28.77, 28.74, 24.6, 24.5, 22.3, 13.5, 8.1; ^{31}P NMR (121 MHz, CDCl$_3$/CD$_3$OD/D$_2$O 35:20:3) δ - ?0.05 ppm; ESI-MS (–ve ion) *m/z* 1102(80), 1101(69); HRMS ESI (-ve) (Found: *m/z* 1101.6702); C$_{60}$H$_{118}$NO$_{20}$P requires *m/z* 1101.6679.
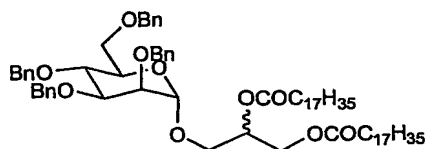
25

EXAMPLE 1(c):   Synthesis of Compound 17

The total synthesis of compound 17 is represented schematically in Figure 3.

95896-1

Preparation of Di-stearate 16

16

A mixture of the diol **8** (127 mg, 0.21 mmol), stearoyl chloride (152 mg, 0.50 mmol) and pyridine (80 μl, 1.00 mmol) in dry dichloromethane (10 ml) was stirred at ambient temperature for 17 h. The mixture was diluted with dichloromethane, washed with 10% hydrochloric acid solution, saturated sodium bicarbonate solution, brine then dried over magnesium sulfate. Removal of the solvent and purification of the residue by silica gel column chromatography gave a 1:1 mixture of the stereoisomeric diglycerides **16** (200 mg, 84%) as a waxy white solid.

Preparation of Mannopyranosyl glyceride 17 (Compound 17)

1,2-Di-*O*-stearoyl-3-α-D-mannopyranosylglycerol
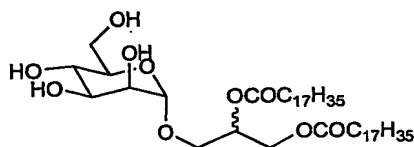
17

A mixture of the glyceride **16** (121 mg, 0.105 mmol) and 10% palladium on carbon (50 mg) in ethanol (25 ml) was stirred under an atmosphere of hydrogen 20 h. The mixture was filtered through a pad of Celite and the filter cake was washed with a mixture of methanol and dichloromethane. The solvent was removed from the filtrate to give a white solid (93 mg) which was subjected to silica gel column chromatography (CH₂Cl₂/MeOH 95:5 to 90:10 gradient elution) to give a 1:1 mixture of the steroisomeric diacyl glyerides **17** (48 mg, 58%) as a white amorphous solid.

EXAMPLE 2: *In vivo* **Efficacy**

Mouse Eosinophilia Model

Model

An ovalbumin (OVA) induced airway eosinophilia mouse model of atopic airway inflammation was used to determine the effectiveness of the synthetic molecules in suppressing the development of airway eosinophilia. This model is widely used to establish "asthma-like effects" in mice – see for example, Erb *et al.,* (1998); Herz *et al.,* (1998); and Randaolf *et al.,* (1999).

Mice

C57B1/6J mice were bred and housed at the Wellington School of Medicine Animal Facility (Wellington, New Zealand). The experimental procedures were approved by the animal ethics committee and were in accordance with University of Otago (Dunedin, New Zealand) guidelines for care of animals.

OVA-induced airway inflammation

OVA Sensitisation – 6 to 8 week-old mice (4 to 5 mice per group) were injected intraperitoneally (i.p.) with 2 µg ovalbumin in 200 µl alum adjuvant at day 0. A booster intraperitoneal injection of 2 µg ovalbumin in 200 µl alum adjuvant was administered at day 14.

Experimental treatments

The mice were randomly allocated to treatment groups with control mice receiving only PBS (Phosphate Buffered Saline), while treated mice received either PIM extracted from *Mycobacterim bovis*, or one of the synthetic molecules.

Treatment protocols with PIM extract or synthetic molecules

7 to 14 days following the second i.p. injection, mice were anaesthetised. Each mouse was treated intranasally as outlined in Table 1 with the indicated concentrations of PIM extract or

95896-1

a synthetic molecule of the invention in 50 µl of PBS. Control mice were given PBS intranasally.

Table 1. Summary of Experiments

Pim Type	Dose Rates (µg/mg)	Number Of Mice
M. Bovis PIM	0, 0.02, 0.2, 2.0	17 including 5 controls*
Compound 15	0, 0.02, 0.2, 2.0	22 including 5 controls*

5   * The same controls (n=5) were used for each treatment group.

OVA challenge – 7 days following treatment with the test molecules, mice were anaesthetised and challenged intranasally with 50 µl of 2 mg/µl ovalbumin in PBS.

10   **Measurements of airway eosinophilia**

4 days after intranasal airway challenge with OVA the mice were sacrificed. The trachea was cannulated and bronchoalveolar lavage (BAL) was performed (3 x 1 ml PBS). Total BAL cell numbers were counted and spun onto glass slides using a cytospin. Percentages of eosinophils, macrophages, lymphocytes and neutrophils were determined microscopically 15   using standing histological criteria.

Results

Table 2. Least square means of eosinophil counts ($\times 10^6$) by dose rate of the test molecule

Dose Rate (µg/ml)	Mean Eosinophil Count ($\times 10^6$)*	s.e.m.
0	0.272^a	0.040
0.02	0.084^b	0.044
0.2	0.123^b	0.044
2	0.114^b	0.042

*Means with different letters are significantly different (P<0.05).

Table 3. Least square means of Eosinophilia counts (*10^6) by test molecule type

Test Molecule Type	Mean Eosinophil Count ($\times 10^6$)	s.e.m
M. bovis PIM	0.153	0.031
Compound 15	0.144	0.030

The results of one comparative experiment are presented in Tables 2 and 3 and Figure 4. The data were analysed as a single group as the experiment tested two molecules at the same time. The statistical model included terms for dose and molecule type as fixed effects and the dose by molecule type interaction. The mean eosinophil counts for the two molecule types are presented in Figure 4.

Overall, the dose of PIM extract or synthetic molecule was highly significant (P<0.01). All animals treated with either PIM extract or a synthetic molecule had a reduced eosinophil count compared to the control animals. No significant differences were found between dose rates in the pair-wise comparisons (Table 2). The overall effect of the type of molecule was not significant (P>0.05; Table 3). Both molecule types appear to be equally effective in reducing the eosinophilia in this experiment meaning that it is likely the active structural element or elements required to produce this effect on the degree of eosinophilia are present in both molecules.

Compound 17 was not biologically active. As this molecule did not include an E or B group, this result indicated that at least one or both such groups are essential for biological activity (results not shown).

EXAMPLE 3:   **Pharmaceutical Formulations**

The compounds suitable for use in the present invention may be administered alone, although it is preferable that they be administered as a pharmaceutical formulation. The compounds of the invention are highly biologically active and it is anticipated that, for airways or nasal mucosal administration, from 1 to 500µg/ml of the active ingredient would be present in the formulation.

95896-1

REFERENCES

Remington's Pharmaceutical Sciences, 16th edition, Oslo, A. (ed), 1980.

5   Erb, K.J., Holloway, J.W., Sobeck, A., Moll, H., Le Gros, G. (1998). Infection of mice with Mycobacterium bovis Calmette-Guerin (BCG) suppresses allergen-induced airway eosinophilia. *J. Exp. Med.* 187: 561-569.

Herz, U., Gerhold, K., Gruber, C., Braun, A., Wahn, U., Renz, H., Paul, K. (1998). BCG infection suppresses allergic sensitisation and development of increased airway reactivity in
10   an animal model. *J. Allergy Clin. Immunol.* 102: 867-874.

Randolf, D.A., Stephens, R., Carruthers, C.J., Chaplin, D.D. (1999).   Cooperation between Th1 and Th2 cells in a murine model of eosinophilic airway inflammation *J. Clinical Investigation*, 104:1021-1029.

15

Crossman, A., Jr., Brimacombe, J. S. & Ferguson, M. A. J. (1997). Parasite glycoconjugates. Part 7. Synthesis of further substrate analogs of early intermediates in the biosynthetic pathway of glycosylphosphatidylinositol membrane anchors. *Journal of the Chemical Society, Perkin Transactions 1: Organic and Bio-Organic Chemistry*, 2769-2774.

20

Gigg, J., Gigg, R., Payne, S. & Conant, R. (1985). The allyl group for protection in carbohydrate chemistry. Part 15. Synthesis of propyl 4-O-(3,6-di-O-methyl-b-D-glucopyranosyl)-2,3-di-O-methyl-a-D-rhamnopyranoside. *Carbohydrate Research* **141**, 91-97.

25

Gilleron, M., Quesniaux, V. F. J. & Puzo, G. (2003). Acylation state of the phosphatidylinositol hexamannosides from Mycobacterium bovis Bacillus Calmette Guerin and Mycobacterium tuberculosis H37Rv and its implication in Toll-like receptor response. *Journal of Biological Chemistry* **278**, 29880-29889.

30

Gilleron, M., Ronet, C., Mempel, M., Monsarrat, B., Gachelin, G. & Puzo, G. (2001). Acylation state of the phosphatidylinositol mannosides from Mycobacterium bovis bacillus Calmette Guerin and ability to induce granuloma and recruit natural killer T cells. *Journal of Biological Chemistry* **276**, 34896-34904.

35

Green, M. M., Gross, R. A., Cook, R. & Schilling, F. C. (1987). Broken worm and worm-like models for polyisocyanates. *Macromolecules* **20**, 2636-2638.

Hirth, G. & Barner, R. (1982). Synthesis of glyceryl ether phosphatides. Part 1. Preparation
40   of 1-O-octadecyl-2-O-acetyl-sn-glyceryl-3-phosphorylcholine ('platelet activating factor'), its enantiomers and some analogous compounds. *Helvetica Chimica Acta* **65**, 1059-1084.

Hirth, G., Saroka, H., Bannwarth, W. & Barner, R. (1983). Synthesis of glyceryletherphosphatides. Part 2. Preparation of 2-O-acetyl-1-O-[(Z)-9-octadecenyl]-sn-glyceryl-3-phosphorylcholine ('Oleyl-PAF'), of its enantiomer and some analogous, unsaturated compounds. *Helvetica Chimica Acta* **66**, 1210-1240.

Joyce, S., & Van Kaer, L. (2003). CD1-restricted antigen presentation:an oily matter. *Current opinion in immunology* **15**, 95-104.

Koizumi, K., Tanimoto, T., Okada, Y., Nakanishi, N., Kato, N., Takagi, Y. & Hashimoto, H. (1991). Characterization of five isomers of branched cyclomaltoheptaose (b CD) having degree of polymerization (d.p.) = 9: reinvestigation of three positional isomers of diglucosyl-b CD. *Carbohydrate Research* **215**, 127-136.

Koto, S., Morishima, N., Miyata, Y. & Zen, S. (1976). Preparation of 2,3,4,6-tetra-O-benzyl-D-mannose. *Bulletin of the Chemical Society of Japan* **49**, 2639-2640.

Koto, S., Morishima, N., Shichi, S., Haigoh, H., Hirooka, M., Okamoto, M., Higuchi, T., Shimizu, K., Hashimoto, Y. & et al. (1992). Dehydrative glycosylation using heptabenzyl derivatives of glucobioses and lactose. *Bulletin of the Chemical Society of Japan* **65**, 3257-3274.

Lindberg, J., Ekeroth, J. & Konradsson, P. (2002). Efficient Synthesis of Phospholipids from Glycidyl Phosphates. *Journal of Organic Chemistry* **67**, 194-199.

Lindhorst, T. K., Dubber, M., Krallmann-Wenzel, U. & Ehlers, S. (2000). Cluster mannosides as inhibitors of type 1 fimbriae-mediated adhesion of Escherichia coli: pentaerythritol derivatives as scaffolds. *European Journal of Organic Chemistry*, 2027-2034.

RajanBabu, T. V., Fukunaga, T. & Reddy, G. S. (1989). Stereochemical control in hex-5-enyl radical cyclizations: from carbohydrates to carbocycles. 3. *Journal of the American Chemical Society* **111**, 1759-1769.

Severn, W.B., Jones, A.M., Kittelberger, R., de Lisle, G.W., and Atkinson, P.H., (1997). Improved procedure for the isolation and purification of lipo-aribomanan Mycobacterium bovis strain AN5. *J. Microbol. Methods* **28**, 123-130.

Watanabe, Y., Nakamoto, C. & Ozaki, S. (1993). Glycosidation based on phosphite chemistry. *Synlett*, 115-116.

Watanabe, Y., Nakamoto, C., Yamamoto, T. & Ozaki, S. (1994). Glycosylation using glycosyl phosphite as a glycosyl donor. *Tetrahedron* **50**, 6523-6536.

INDUSTRIAL APPLICATION

As will be appreciated from the above, the primary application of the invention is in the treatment of inflammatory or immune cell-mediated disorders. That treatment may be
5   prophylactic, to prevent risk of developing such diseases or disorders, or therapeutic, to suppress established disease or symptoms.

The pharmaceutical compositions of the invention may be formulated for administration by any route depending on the nature of the disease or disorder to be treated. For example, for
10   asthma, the composition will preferably be formulated for respiratory administration by the intranasal or inhaled route. For arthritis or diabetes, the composition will preferably be formulated for oral or subcutaneous administration. For rhinitis, the compositions will preferably be formulated for oral, or nasal mucosal delivery.

15   The invention also provides a process for efficient and economical large scale production of the synthetic molecules of the invention for use as an active ingredient in the pharmaceutical compositions.

It will be appreciated that the present invention is not limited to the above examples only,
20   many variations, which may readily occur to a skilled worker, being possible without departing from the scope of the invention.
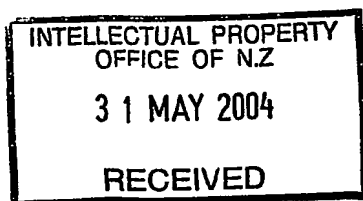
The Malaghan Institute of
Medical Research University
of Otago and AgResearch
by the authorised agents

per *[signature]*

95896-1

Document made available under the Patent Cooperation Treaty (PCT)

International application number: PCT/NZ04/000293

International filing date: 18 November 2004 (18.11.2004)

Document type: Certified copy of priority document

Document details: Country/Office: NZ
Number: 533245
Filing date: 31 May 2004 (31.05.2004)

Date of receipt at the International Bureau: 17 January 2005 (17.01.2005)

Remark: Priority document submitted or transmitted to the International Bureau in compliance with Rule 17.1(a) or (b)

World Intellectual Property Organization (WIPO) - Geneva, Switzerland
Organisation Mondiale de la Propriété Intellectuelle (OMPI) - Genève, Suisse